

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

Cord Blood Metabolic Signatures of Birth Weight: A Population-Based Study

This is the author's manuscript

Original Citation:

Availability:

This version is available <http://hdl.handle.net/2318/1663078> since 2020-04-05T10:40:37Z

Published version:

DOI:10.1021/acs.jproteome.7b00846

Terms of use:

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

Cord Blood Metabolic Signatures of Birth Weight: A Population-Based Study

Robinson O, Keski-Rahkonen P, Chatzi L, Kogevinas M, Nawrot T, Pizzi C, Plusquin M, Richiardi L, Robinot N, Sunyer J, Vermeulen R, Vrijheid M, Vineis P, Scalbert A, Chadeau-Hyam M

Abstract

Birth weight is an important indicator of maternal and fetal health and a predictor of health in later life. However, the determinants of variance in birth weight are still poorly understood. We aimed to identify the biological pathways, which may be perturbed by environmental exposures, that are important in determining birth weight. We applied untargeted mass-spectrometry-based metabolomics to 481 cord blood samples collected at delivery in four birth cohorts from across Europe: ENVIRONAGE (Belgium), INMA (Spain), Piccolipiu (Italy), and Rhea (Greece). We performed a metabolome-wide association scan for birth weight on over 4000 metabolic features, controlling the false discovery rate at 5%. Annotation of compounds was conducted through reference to authentic standards. We identified 68 metabolites significantly associated with birth weight, including vitamin A, progesterone, docosahexaenoic acid, indolelactic acid, and multiple acylcarnitines and phosphatidylcholines. We observed enrichment ($p < 0.05$) of the tryptophan metabolism, prostaglandin formation, C21-steroid hormone signaling, carnitine shuttle, and glycerophospholipid metabolism pathways. Vitamin A was associated with both maternal smoking and birth weight, suggesting a mediation pathway. Our findings shed new light on the pathways central to fetal growth and will have implications for antenatal and perinatal care and potentially for health in later life.

Keywords:

birth weight; cord blood; fetal growth; metabolism; metabolomics; pathway perturbation

Introduction

Weight at birth is of importance due to its relation to the health of both the mother and the newborn⁽¹⁾ and is a predictor of the subsequent development of the child.^(2, 3) Low birth weight has also been associated with disease in later life, including cardiovascular disease and diabetes⁽⁴⁾ while high birth weight is associated with increased risk of developing breast cancer and other malignancies.^(5, 6) Thus under the developmental origin of health and disease hypothesis, adverse fetal development may have a lifelong impact. Exposure to a number of different agents during pregnancy including smoking,⁽⁷⁾ air pollution,⁽⁸⁾ and chemicals such as polychlorinated biphenyls⁽⁹⁾ have been associated with lower birth weight. Improved understanding of the biological pathways associated with environmentally induced alterations in birth weight may identify mechanisms through which fetal growth is affected. This may, in turn, both inform primary care and ultimately improve the causal evidence regarding adverse fetal development.

Metabolomics is increasingly used in maternal–fetal medicine⁽¹⁰⁾ to identify biological changes associated with fetal growth. Horgan et al.⁽¹¹⁾ examined ultra-high-performance liquid chromatography–mass spectrometry (UHPLC–MS) profiles of first-trimester maternal plasma samples to identify metabolites predictive of small-for-gestational-age babies. Maitre et al. used nuclear magnetic resonance (NMR) spectroscopy on maternal pregnancy urine samples to identify metabolites predictive of preterm birth,

small-for-gestational-age, and fetal growth restriction(12) and birth weight.(13) Dessì et al.(14) identified four metabolites associated with fetal growth restriction in neonatal urine samples. However, only a handful of studies have investigated metabolic changes in cord blood, which is a particularly relevant tissue because it contains the essential nutrients, hormones, and immunological factors and potentially harmful xenobiotic metabolites, to which the developing fetus is directly exposed. Horgan et al.(11) compared six small-for-gestational-age babies with controls observing differences in levels of sphingolipids, phospholipids, and carnitines. Ivorra et al.(15) and Tea et al.(16) both used NMR spectroscopy to compare a small number of low-birth-weight and very low-birth-weight newborns, respectively, with controls and detected some differences in metabolite levels. Recently, Hellmuth et al.(17) applied a targeted mass-spectrometry-based analysis and observed a positive association between lysophosphatidylcholines (lysoPCs) and birth weight. While initial studies have so far been based on small samples or limited subsets of molecules, they demonstrate the potential of metabolic profiling to detect biological pathways related to fetal development. Adequately powered studies using sensitive and untargeted platforms are now required.

In this study, we have employed untargeted UHPLC–MS-based metabolomics to identify metabolic features associated with birth weight in cord blood collected from a large population-based sample from four European birth cohorts. We aimed to understand the mechanisms that are important to fetal growth, which may be influenced by the maternal environment including exposure to air pollutants and tobacco smoke, and that may impact health over the life course.

Experimental Section

Cohorts and Sampling

Within the context of the EXPOsOMICS collaborative European project,(18) metabolomic analyses were conducted on umbilical cord blood samples from four population-based birth cohorts: ENVIRONAGE,(19) Piccolipiu,(20) INMA,(21) and Rhea.(22) The ENVIRONAGE cohort recruited women when they arrived for delivery at the South-East-Limburg Hospital in Gent, Belgium between 2010 and 2013. The INMA cohort enrolled women during the first trimester of pregnancy at public primary health care centers or hospitals in Sabadell, Spain between 2004 and 2006. The participating Piccolipiu cohort center enrolled women giving birth at the main hospital of the City of Turin, Italy, between 2011 and 2013. The Rhea cohort enrolled women during the first trimester of pregnancy at public primary health care centers or hospitals in Heraklion, Greece between 2007 and 2008. Whole blood samples were collected using venipuncture of cord vessels before the placenta was delivered and processed as follows in each cohort: In ENVIRONAGE, samples were collected into EDTA (BD, Franklin Lakes, NJ) vacutainers and within 20 min were centrifuged at 3200 rpm for 15 min into plasma. In Piccolipiu, samples were collected into BD EDTA vacutainers, stored at 4 °C for <24 h, and centrifuged for 10 min at 1300g into plasma. In Rhea, samples were collected into BD gel separator vacutainers and centrifuged within 2 h at 2500 rpm for 10 min into serum. In INMA, samples were collected into BD gel separator vacutainers, stored at 4 °C for <4 h, and centrifuged at 3000 rpm for 10–15 min into serum. Samples were immediately frozen at –20 °C (INMA) or –80 °C (all other cohorts) until analysis. Cohort inclusion criteria and further protocols can be found in the respective cohort references.

Family lifestyle factors were collected from mothers through interview by trained fieldworkers and medical history for each family transferred from hospital records. Samples were selected from each cohort on the basis of biomaterial and data availability. Selected samples were shipped to International Agency for Research on Cancer, Lyon, France for metabolomics analysis.

Exposure Assessment

Exposure to air pollutants (particulate matter $\leq 10 \mu\text{m}$ (PM_{10}), $\leq 2.5 \mu\text{m}$ ($\text{PM}_{2.5}$) and $\leq 0.1 \mu\text{m}$ (UFP) and NO_2) was assessed at the home address, averaged over the year before pregnancy, by land use regression models.(23)

Sample Analysis

Samples were randomized and prepared by mixing a $30 \mu\text{L}$ aliquot with $200 \mu\text{L}$ of acetonitrile and filtering the precipitate with $0.2 \mu\text{m}$ Captiva ND plates (Agilent Technologies). The filtrate was collected into a polypropylene well plate that was sealed and kept refrigerated until analysis. A quality control (QC) sample was prepared by mixing small aliquots of 91 randomly selected study samples. Aliquots ($30 \mu\text{L}$) of the QC sample were then processed along with the study samples, with each 96-well plate containing four individually extracted QCs. Samples were analyzed as a single uninterrupted batch with a UHPLC–MS system consisting of a 1290 Binary LC system, a Jet Stream electrospray ionization (ESI) source, and a 6550 QTOF mass spectrometer (Agilent Technologies). The autosampler tray was kept refrigerated, and $2 \mu\text{L}$ of the sample solution was injected on an ACQUITY UPLC HSS T3 column ($2.1 \times 100 \text{ mm}$, $1.8 \mu\text{m}$; Waters). Column temperature was 45°C and mobile phase flow rate was 0.4 mL/min , consisting of ultrapure water and LC–MS-grade methanol, both containing 0.05% (v/v) of formic acid. The gradient profile was as follows: 0–6 min: $5\% \rightarrow 100\%$ methanol, 6–10.5 min: 100% methanol, 10.5–13 min: 5% methanol. The mass spectrometer was operated in positive polarity using the following conditions: drying gas (nitrogen) temperature 175°C and flow 12 L/min , sheath gas temperature 350°C and flow 11 L/min , nebulizer pressure 45 psi , capillary voltage 3500 V , nozzle voltage 300 V , and fragmentor voltage 175 V . Data were acquired in centroid format using an extended dynamic range mode, with a scan rate of 1.67 Hz and a mass range from 50 to 1000. For MS/MS analyses the isolation width was 1.3 Da and collision energies were 10, 20, and 40 V . Continuous mass axis calibration was performed using two reference ions (m/z 121.050873 and m/z 922.009798). The analytical run was initiated with priming injections of in-house human plasma extract to achieve a stable response, followed by the study samples and one QC sample after every 12 injections. Data were acquired using MassHunter Acquisition B.05.01 software.

Data Preprocessing

Preprocessing of the acquired data was performed using Qualitative Analysis B.06.00 SP1, DA Reprocessor, and Mass Profiler Professional 12.1 software (Agilent Technologies). The initial processing was performed with Molecular Feature Extraction (MFE) algorithm for small molecules using a mass range of 50–1000. Thresholds for the mass and chromatographic peak heights were 1500 and 10 000 counts, respectively. Quality score threshold was 80. Only singly charged proton adducts ($[\text{M}+\text{H}]^+$) were included. Spacing tolerance for isotope peaks was $0.0025 m/z$ plus 7 ppm. The isotope model for common organic molecules was used, and features with indeterminate neutral mass were excluded. Feature alignment was performed with retention time and mass windows of 0.075 min and $15 \text{ ppm} + 2 \text{ mDa}$. A target list for a recursive extraction was created from features present in at least 2% of the samples. Find by Formula (FBF) algorithm was then employed with match tolerance for the mass and retention time $\pm 10 \text{ ppm}$ and $\pm 0.04 \text{ min}$, respectively. Ion species was limited to $[\text{M}+\text{H}]^+$, and a threshold for chromatographic peak height was 2000 counts. The resulting features were aligned in Agilent Mass Profiler Professional using the same parameters as described above. For statistical analysis, metabolic features present in $<60\%$ of the samples were removed, data were log-transformed, and missing values were imputed using the impute.QRILC function within the imputeLCMD R package.(24)

Statistical Analysis

The relationship between birth weight and the cord metabolome was assessed using a metabolome-wide association scan (MWAS) approach with separate linear regression models for each metabolic feature using the “omics” R package.(25) To account for multiple comparisons, we applied a Benjamini–Hochberg correction using an overall false discovery rate of (FDR) <5%. The covariates included in the main MWAS analysis were gestational age, sex, cohort, maternal height, maternal weight, and paternal height. Covariates were chosen first following visualization of assumptions using a directed acyclical graph (Figure s1) and then following testing of associations with both birth weight in bivariate analyses (Analysis of Variance or Pearson’s correlation tests) and with metabolic features as visualized by Q–Q plots of the p -value distribution. We chose to not adjust for environmental factors that may be associated with birth weight in the main analysis because we hypothesized that the metabolome may mediate these associations. Instead we tested for potential confounding by environmental factors in sensitivity analyses through stratification by covariates and adjustment separately for each birth-weight-associated risk factor.

We further investigated links between risk factors and birth weight by first constructing a linear regression model of birth weight, including those factors associated with birth weight ($p \leq 0.1$) in bivariate analyses. We then adjusted the model for the metabolome, using the first components of a principal component analysis (PCA) of metabolites associated with birth weight in the main MWAS analysis. The number of components to include was selected by examination of a scree plot of explained variance. Where risk factor associations were attenuated following adjustment on birth-weight-related metabolites, we tested the association of that factor with all birth-weight-related metabolites in a further analysis.

All analyses were performed in R version 3.3.(26)

Metabolite and Pathway Annotation

Annotation of the discriminating features was done as follows: (1) The m/z values of all features were searched against the human metabolite database(27) and Metlin(28) using $[M+H]^+$, $[M+Na]^+$, and $[M+2H]^{2+}$ as adducts and ± 8 ppm for molecular-weight tolerance. In addition, MyCompoundID metabolite library(29) was searched for potential conjugates (sulfates, glucuronides) and neutral losses ($-NH_3$, H_2O) using $[M+H]^+$ ions and ± 8 ppm mass tolerance. (2) Features were grouped based on retention time similarity and intensity correlation across the samples to assist in identifying ions originating from the same metabolite. (3) Quality of the chromatographic peaks and spectra were inspected, and the plausibility of database candidates was assessed based on retention time, isotope pattern, and adduct formation or neutral losses. (4) Identification was confirmed by reanalysis of representative samples and pure standards when available and comparison of the retention times and the MS/MS spectra. When standards were not available, MS/MS spectra were acquired and compared against those in mzCloud (www.mzcloud.org) or Metlin. Chromatograms and spectra can be found in the Supporting Information (Additional Data Set 1). The level of identification was as proposed by Sumner et al.(30)

Significantly enriched metabolic pathways were identified using the *Mummichog* program.(31) The algorithm searches tentative compound lists from metabolite reference databases against an integrated model of human metabolism to identify functional activity. Fisher’s exact tests and permutation are used to infer p values for likelihood of pathway enrichment among significant features as compared with pathways identified among the entire compound set present in reference list (the entire metabolome data set), considering the probability of mapping the significant m/z features to pathways. *Mummichog* parameters were set to match against ions produced by the MS method employed ($[M]^+$, $[M+H]^+$, $[M+2H]^{2+}$, $[M+3H]^{3+}$,

[M+Na]⁺, [M+H+Na]²⁺, [M+K]⁺, [M-H₂O+H]⁺, [M-H₄O₂+H]⁺, [M-NH₃+H]⁺, [M-CO+H]⁺, [M-CO₂+H]⁺, [M-HCOOH+H]⁺, [M+HCOONa]⁺, [M-HCOONa+H]⁺, and [M-C₃H₄O₂+H]⁺) at ±8 ppm mass tolerance.

Results

Participant Information

Samples of cord blood collected from 499 deliveries (200 from ENVIRONAGE, 100 each from INMA and Rhea, and 99 from Piccolipiu) were included in the analysis. The mean birth weight was 3309 g (interquartile range 2992–3598 g) with 16 (3%) babies born with low birth weight (<2500g). Demographic information and covariate associations with birth weight are shown in Table 1. Gestational age, cohort, sex, maternal height, weight and body mass index, paternal height, maternal weight gain during pregnancy, smoking by the mother, and residential exposure to air pollution were all significantly associated with birth weight in bivariate analyses.

Table 1. Participant Information and Covariate Associations with Birth Weight

	<i>N</i> (%) or mean (IQR)	missing values	mean birth weight (IQR) or <i>r</i> with birth weight	<i>p</i> value ^a
birth weight (g)	3309 (2992–3598)	1	–	–
gestational age (weeks)	39.2 (38.1–40)	1	0.21	<0.0001
cohort		0		0.014
Rhea	100 (20)	–	3220 (2970–3520)	
Environage	200 (40)	–	3384 (3045–3705)	
Piccolipiu	99 (20)	–	3221 (2955–3490)	
Inma	100 (20)	–	3298 (2998–3572)	
sex		1		0.0005
male	258 (52)	–	3377 (3076–3648)	
female	240 (48)	–	3237 (2915–3550)	
parity		4		0.13
0	129 (26)	–	3275 (2990–3570)	
1	238 (48)	–	3294 (2971–3579)	
2	128 (260)	–	3379 (3119–3640)	
season of conception		3		0.35
January–March	122 (25)	–	3318 (3000–3550)	
April–June	129 (26)	–	3296 (2970–3600)	
July–September	160 (32)	–	3348 (3019–3630)	

	<i>N</i> (%) or mean (IQR)	missing values	mean birth weight (IQR) or <i>r</i> with birth weight	<i>p</i> value ^a
October–December	85 (17)	–	3240 (2980–3485)	
delivery		2		0.18
vaginal	382 (77)	–	3324 (3000–3614)	
caesarean	115 (23)	–	3259 (2960–3542)	
maternal age (y)	30.7 (27.9–34)		–0.04	0.44
maternal height (cm)	164.5 (160–168)	2	0.27	<0.0001
maternal weight (kg)	62 (42–130)	2	0.24	<0.0001
maternal BMI	23 (20.8–25.8)	2	0.15	0.001
maternal weight gain (kg)	13.63 (10–17)	13	0.21	0.0002
paternal height (cm)	177.8 (173–182)	17	0.14	0.003
paternal weight (kg)	82 (73–90)	17	0.07	0.11
mother born in country		9		0.69
yes	432 (88)	–	3315 (3000–3586)	
no	58 (12)	–	3287 (2881–3638)	
mother's education		12		0.64
primary school	62 (13)	–	3291 (2975–3595)	
secondary school	209 (43)	–	3296 (2980–3590)	
university or higher	216 (44)	–	3333 (3060–3589)	
father's education		28		0.08
primary school	96 (20)	–	3236 (2915–3550)	
secondary school	227 (47)	–	3288 (2982–3580)	
university or higher	148 (30)	–	3365 (3084–3600)	
maternal smoking (during second trimester)		3		0.06
yes	57 (11%)	–	3207 (2890–3440)	
no	439 (89%)	–	3324 (3000–3612)	
passive smoke exposure		15		0.50
yes	172 (36)	–	3295 (2970–3616)	
no	312 (64)	–	3324 (3000–3592)	

	<i>N</i> (%) or mean (IQR) missing values	mean birth weight (IQR) or <i>r</i> with birth weight	<i>p</i> value ^a
exposure to PM ₁₀ (µg/m ³)	33.0 (17.8–44.0) 4	–0.13	0.002
exposure to PM _{2.5} (µg/m ³)	18.14 (12.4–21.0) 4	–0.11	0.02
exposure to NO ₂ (µg/m ³)	29.1 (13.4–40.3) 4	–0.11	0.02

a

p value for association with birth weight, calculated from Pearson's correlation test (continuous variable) or analysis of variance test (categorical variable). IQR = Interquartile range.

Metabolomic Data

The total number of metabolomic features was 9947. Out of these, 4870 (49%) were found in at least 90% of the QC sample injections, with 4019 (83%) having a coefficient of variation <30%, indicating good reproducibility. Features were excluded if present in <60% of participant samples, leaving 4714 metabolic features for statistical analysis. A PCA of these features (whole metabolome PCA) showed that the first component explained 18% of variance and 307 components explained 95% of the variance, suggesting considerable redundancy within the data (Figure s2 (top)). There was clear separation along the second component by cohort (Figure s2 (bottom)). No separation was observed with other covariates or technical variables.

Metabolic Features Associated with Birth Weight

In the main MWAS analysis of 481 participants with complete covariate data, adjusted for gestational age, cohort, sex, maternal height, maternal weight, and paternal height, 138 features were significantly associated (FDR < 5%) with birth weight (Figure 1).

After grouping of ions originating from the same molecule (matched by retention time and pairwise feature correlation), there were 68 unique compounds associated with birth weight. Twenty-nine of these were annotated to at least the level of compound class based on matching mass, isotope pattern, and product ion spectra (Table 2). Fifteen compounds were identified as acylcarnitines, nine as phosphatidylcholines (PCs) or lysoPCs, two as tryptophan metabolites (indolelactic acid and an isomeric form of methoxykynurenate), two as essential nutrients (retinol (vitamin A) and docosahexaenoic acid (DHA)), and one as a steroid hormone. Seven of these were conclusively identified through comparisons with authentic standards: decanoylcarnitine, dodecanoylcarnitine and tetradecanoylcarnitine, indolelactic acid, retinol, the ω-3 fatty acid DHA, and progesterone, one of the major progestational steroid hormones. In addition to the 29 annotated metabolites, five compounds (three diacylglycerols and two lysoPCs) were tentatively assigned based on exact mass and isotope pattern due to low intensity or lack of MS/MS spectra or authentic standards (Table 2). Retention time and exact mass of all significantly associated features, including unassigned compounds, is given in Table s1. Chromatograms and mass spectra of all annotated compounds are given in the Supporting Information in Additional Data Set 1.

Table 2. Metabolites Significantly Associated with Birth Weight

<i>m/z</i>	retention time (min)	ion	name ^a	ID level ^b	compound class	direction of change with increasing birth weight	<i>P</i> value
368.2793	5.631	[M+H] ⁺	3,5-tetradecadienylcarnitine (C14:2)	2	acylcarnitine	DOWN	1.00 × 10 ^{−8}
518.3216	6.782	[M+H] ⁺	LysoPC (C18:3)	2	glycerophospholipid	UP	4.63 × 10 ^{−8}

<i>m/z</i>	retention time (min)	ion	name ^a	ID level ^b	compound class	direction of change with increasing birth weight	<i>P</i> value
494.3250	6.817	[M+H] ⁺	LysoPC (C16:1)	2	glycerophospholipid	UP	7.45×10^{-8}
344.2797	5.647	[M+H] ⁺	dodecanoylcarnitine (C12:0)	1	acylcarnitine	DOWN	8.31×10^{-8}
206.0822	3.829	[M+H] ⁺	indolelactic acid	1	tryptophan metabolite	DOWN	9.16×10^{-8}
522.3555	6.980	[M+H] ⁺	LysoPC (C18:1)	2	glycerophospholipid	UP	1.45×10^{-7}
641.5112	9.938	[M+Na] ⁺	diacylglycerol (C36:3)	4	glycerolipid	DOWN	2.34×10^{-7}
269.2278	7.219	[M- H ₂ O+H] ⁺	retinol	1	vitamin	UP	1.30×10^{-6}
639.4946	9.408	[M+Na] ⁺	diacylglycerol (C36:4)	4	glycerolipid	DOWN	1.74×10^{-6}
396.3100	5.944	[M+H] ⁺	9,12-hexadecadienoylcarnitine (C14:1)	2	acylcarnitine	DOWN	2.32×10^{-6}
522.3565	7.058	[M+H] ⁺	LysoPC (C18:1)	2	glycerophospholipid	UP	4.55×10^{-6}
329.2482	7.232	[M+H] ⁺	docosaheptaenoic acid	1	fatty acid	DOWN	1.18×10^{-5}
766.5815	8.859	[M+H] ⁺	PC-O (C36:4) (C ₄₄ H ₈₀ NO ₇ P)	3	glycerophospholipid	DOWN	1.20×10^{-5}
316.2489	5.139	[M+H] ⁺	decanoylcarnitine (C10:0)	1	acylcarnitine	DOWN	1.30×10^{-5}
810.6053	9.169	[M+H] ⁺	PC (C38:4)	3	glycerophospholipid	DOWN	1.37×10^{-5}
258.1699	2.831	[M+H] ⁺	2-hexenoylcarnitine (C6:1)	2	acylcarnitine	DOWN	4.43×10^{-5}
342.2641	5.422	[M+H] ⁺	<i>trans</i> -2-dodecenoylcarnitine (C12:1)	2	acylcarnitine	DOWN	4.89×10^{-5}
314.2318	4.967	[M+H] ⁺	decenoylcarnitine_2 (C10:1) ^c	2	acylcarnitine	DOWN	5.20×10^{-5}
370.2955	5.840	[M+H] ⁺	tetradecenoylcarnitine (C14:1)	2	acylcarnitine	DOWN	5.49×10^{-5}
615.4959	9.763	[M+Na] ⁺	diacylglycerol (C34:2)	4	glycerolipid	DOWN	6.82×10^{-5}
734.5700	8.960	[M+H] ⁺	PC (C32:0)	3	glycerophospholipid	DOWN	1.17×10^{-4}
314.2321	4.878	[M+H] ⁺	decenoylcarnitine_1 (C10:1) ^c	2	acylcarnitine	DOWN	1.55×10^{-4}
315.2320	6.394	[M+H] ⁺	progesterone	1	steroid hormone	DOWN	2.11×10^{-4}
288.2171	4.422	[M+H] ⁺	l-octanoylcarnitine (C8:0)	2	acylcarnitine	DOWN	3.21×10^{-4}
232.1537	1.927	[M+H] ⁺	butyrylcarnitine/isobutyryl-l-carnitine (C4:0)	2	acylcarnitine	DOWN	3.61×10^{-4}
220.0605	3.671	[M+H] ⁺	methoxykynurenate (C ₁₁ H ₉ NO ₄) ^d	3	tryptophan metabolite	DOWN	4.69×10^{-4}
548.3681	7.141	[M+H] ⁺	LysoPC (C20:2)	4	glycerophospholipid	UP	5.58×10^{-4}
782.5712	8.628	[M+H] ⁺	PC (C36:4)	3	glycerophospholipid	DOWN	6.03×10^{-4}
372.3112	6.006	[M+H] ⁺	tetradecanoylcarnitine (C14:0)	1	acylcarnitine	DOWN	6.46×10^{-4}
758.5747	8.684	[M+H] ⁺	PC (C34:2)	3	glycerophospholipid	DOWN	7.88×10^{-4}
570.3551	7.021	[M+H] ⁺	LysoPC (C22:5)	4	glycerophospholipid	UP	8.01×10^{-4}
386.2899	5.568	[M+H] ⁺	3-hydroxy- <i>cis</i> -5-tetradecenoylcarnitine (C14:1)	2	acylcarnitine	DOWN	8.10×10^{-4}
412.3045	5.750	[M+H] ⁺	3-hydroxyhexadecadienoylcarnitine (C16:1)	2	acylcarnitine	DOWN	9.82×10^{-4}
398.3264	6.109	[M+H] ⁺	hexadecenoylcarnitine (C16:1)	2	acylcarnitine	DOWN	1.01×10^{-3}

Common names as used in HMDB. Molecular formulas given when identification inconclusive or more than one isomers are possible. Chain lengths and number of double bonds are indicated for acylcarnitines, glycerophospholipid, and diacylglycerols.

Level 1 (identified compounds): retention time and MS/MS spectra matches with an authentic chemical standard; Level 2 (putatively annotated compound): no standard available or analyzed but has a single database candidate within 5 ppm mass error, matching isotope pattern, and MS/MS spectra; Level 3 (putatively characterized compound class): MS/MS spectral similarity with compounds from a known chemical class; Level 4 (unknown compounds): no standard or MS/MS spectra available but a chemically plausible hit in a metabolite database within 5 ppm mass error and a matching isotope pattern.(60)

Two isomers of decenoylcarnitine were identified.

Likely a positional isomer of methoxykynurenate based on closeness of retention time and similarities in spectra to those acquired from authentic standard of 8-methoxykynurenate.

As shown in clustered correlation heatmap (Figure 2), the longer chained acylcarnitines clustered together and were moderately correlated with progesterone, the tryptophan metabolites, and the shorter chained acylcarnitines. The PCs, smaller-sized lysoPCs, and tentatively assigned diacylglycerols formed three further clusters. The diacylglycerol cluster was moderately correlated with DHA. Both the diacylglycerol cluster and DHA were negatively correlated with the cluster of smaller sized lysoPCs.

In PCA of the significant compounds, >50% of variance was explained by 5 components, and 41 components were required to explain >95% of the variance of these features. There was some separation in results by cohort (Figure 3A), and inspection of the component loadings (Figure 3B) revealed this was driven mainly by differing levels of acylcarnitines along the first component and of lysoPCs and the tentatively assigned diacylglycerols along the second component.

Sensitivity Analysis

To further assess the potential cohort-specific results, we also ran our models on each cohort separately (Table s2). Additionally, we stratified by sex and reran analyses after excluding preterm and caesarean deliveries (Table s2). Results were generally consistent across cohorts, although we noted stronger associations for the smaller sized lysoPCs in the INMA cohort and nonsignificant and opposite directions of association for some acylcarnitines (dodecanoylcarnitine, decanoylcarnitine, *trans*-2-dodecenoylcarnitine, and tetradecenoylcarnitine) in Rhea and for the three tentatively assigned diacylglycerols in Piccolipiu. These differences largely reflect the heterogeneity of the metabolic profiles we observed in each cohort (Figure 3). Upon stratification by sex, we observed stronger associations for decanoylcarnitine, tetradecenoylcarnitine, and hexadecenoylcarnitine in girls and a stronger association with PC (C₄₀H₈₀NO₈P) in boys. Associations were similar when preterm and caesarean deliveries were excluded, except for slightly stronger associations with decanoylcarnitine and dodecanoylcarnitine in the analysis of vaginal deliveries only.

To test for the potential confounding of our results we additionally adjusted the main model separately by factors that influence birth weight. Additional factors included exposure to PM₁₀, paternal education, smoking continuing into the second trimester, and maternal weight gain. We also checked for the confounding effects of mode of delivery and cohort differences in the metabolome data, using the second component of the whole metabolome PCA. As shown in Table s3, associations were robust to adjustment by these factors, indicating negligible confounding.

To investigate the role of the metabolome in potentially mediating the effect of risk factors on birth weight, we adjusted a risk factor model of birth weight (including exposure to PM₁₀, maternal and paternal education, smoking continuing into the second trimester, and maternal weight gain) by the first five principal components of birth-weight-associated metabolites (explaining 50.2% of the total variance in the set of 68 metabolites). Of the included risk factors, only the effect size estimate linking birth weight and smoking continuing into the second trimester was decreased upon adjustment on birth-weight-related metabolites (Table 3). Specifically, while the unadjusted model estimated that babies of mothers who smoked during the second trimester weighed 88.5 g (95% confidence interval (CI): −197.6 g, 20.5 g) less at birth than babies of mothers who did not, that weight loss dropped to 74.0 g (95% CI: −170.7 g, 22.6 g) in the model adjusting for birth-weight-related metabolites.

Table 3. Associations of Risk Factors with Birth Weight, with and without Adjustment for Metabolites

	risk factor model ^a	risk factor model, adjusted for birth-weight-associated metabolome ^b	risk factor model, adjusted for retinol levels in cord blood ^c
	β (95% CI)	β (95% CI)	β (95% CI)
exposure to PM ₁₀ during pregnancy	−3.37 (−11.74, 5.00)	−4.97 (−12.40, 2.45)	−2.90 (−11.09, 5.28)
education level of father (ref primary school)			
secondary school	59.7 (−29.56, 148.96)	80.83 (2.11, 159.54)	71.77 (−15.65, 159.19)
university or higher	116.08 (16.18, 215.98)	126.04 (38.11, 213.97)	119.19 (21.52, 216.86)
mother smoked during second trimester	− 88.54 (−197.62, 20.54)	−74.02 (−170.69, 22.65)	−45.06 (−153.30, 63.18)
maternal weight gain	12.42 (5.77, 19.07)	13.87 (8.00, 19.75)	11.69 (5.18, 18.19)

Adjusted for gestational age, cohort, maternal height and prepregnancy weight, sex of baby, and paternal height.

Adjusted for gestational age, cohort, maternal height and prepregnancy weight, sex of baby, and paternal height and first five principal components of PCA on all birth-weight-associated metabolites.

Adjusted for gestational age, cohort, maternal height and prepregnancy weight, sex of baby, and paternal height and levels of retinol in cord blood.

Linear models investigating the association between the birth-weight-associated metabolites and maternal smoking, adjusted for cohort, maternal education, and body mass index, showed that only blood retinol levels were significantly different (lower, $p = 4.26 \times 10^{-4}$) among mothers who smoked during the second trimester (Figure 4). On the basis of this observation, we additionally looked at the evolution of association between birth weight risk factors in a model adjusting for retinol levels (Table 3). The effect size estimate of smoking on birth weight was halved upon adjustment of retinol levels ($\beta = -45.1$ g, 95% CI: −153.3 g, 63.2 g). Although these changes are not statistically significant, they indicate that retinol may partly mediate the effects of smoking on birth weight.

Pathway Analysis

Mummichog software assigned tentative annotations to 1629 of the 4714 features analyzed and assigned tentative annotations to 30 of the 138 features associated with birth weight (Supporting Information, Additional Data Set 2). Figure 5 shows significantly enriched pathways ($p < 0.05$) identified using *Mummichog*. We observed the enrichment of eight different pathways; two of these involve the synthesis and metabolism of signaling molecules, prostaglandins, and steroid hormones, known to be involved in embryogenesis and child birth.

Discussion

This is the largest study to date investigating untargeted metabolic profiles of cord blood associated with birth-related outcomes, including weight at birth. Variance in birth weight, a trait that reflects the in uterine conditions throughout pregnancy and is an important predictor of health later in life, is determined by a complex combination of factors. Twin studies demonstrated the majority of these factors are of environmental origin.(32) Here we have shown a metabolic signature associated with birth weight, after controlling for hereditary factors such as parental size, among healthy deliveries from the general European population. We observed changes in levels of vitamin A, progesterone, and molecules involved in pathways related to tryptophan metabolism, carnitine shuttle, fatty acid, and glycerophospholipid metabolism.

Levels of vitamin A were higher with higher birth weight, confirming findings from previous studies.(33-36) Vitamin A likely promotes fetal growth through its role in cell proliferation and embryogenesis and interaction with nuclear receptors to alter gene expression.(37) While our results are consistent with changes in cord blood levels of vitamin A associated with smoking previously reported,(38) this is the first study to the best of our knowledge that has demonstrated a link between vitamin A and both smoking and birth weight in the same population. Cigarette smoke has been shown to induce vitamin A depletion in animal models and has been proposed to result from the induction of the CYP1A1 and CYP1A2 enzymes and the subsequent increase in catabolism of retinoic acid.(39) Vitamin A depletion, therefore, may present an important etiological pathway linking smoking with lower birth weight, which should be further investigated.

Progesterone levels were down-regulated with increasing birth weight. This corroborates findings from a recent targeted study of hormone levels in cord blood among Chinese and American births.(40) Progesterone is the major progestational hormone involved throughout all stages of pregnancy. It is required for implantation, maintaining intrauterine conditions, and initiating the signaling cascade to induce labor. Evidence of the relationship of progesterone measured earlier in pregnancy and birth weight is inconsistent.(41-43) However, levels of progesterone in cord blood appear unrelated to levels measured in maternal plasma earlier in pregnancy.(44) This reflects the transition in endocrine metabolism during fetal development, with the fetal endocrine system being well developed by late pregnancy. Pregnenolone, the precursor in the production of progesterone by the placenta, is transported to the fetus for metabolism of large quantities of C-19 steroid hormones by the fetal adrenal glands, which, in turn, signal back to regulate hormonal production by the maternal and placental systems.(45) The size of the fetus may influence the production of molecules such as progesterone around the time of labor onset, for instance, to increase muscle contractibility during delivery. Furthermore, increased activity of the fetal endocrine system in larger babies may reduce the availability of pregnenolone for progesterone production.

We observed a negative association with birth weight and methoxykynurenate and indolelactic acid. These metabolites, never previously associated with birth weight, are both final products of tryptophan metabolism, albeit through different routes, the kynurenine and indole pathways. Tryptophan itself was identified among the annotated features, and although there was no statistically significant association between the amino acid and birth weight ($p = 0.07$), our results indicate altered utilization of tryptophan. Progesterone has been demonstrated to regulate tryptophan metabolism through the inhibition of Trp 2,3-dioxygenase.(46) Tryptophan is essential throughout pregnancy, first to meet the demand for protein synthesis during fetal development, to meet serotonin and kynurenine requirements, and to ultimately

provide quinolinate for the production of NAD^+ , which plays a key role in mitochondrial function.(47) All of these requirements will increase with greater fetal growth, thereby leaving less free tryptophan for conversion into methoxykynurenate and indolelactic acid. Similar results were reported by Favretto et al.,(48) who observed higher levels of tryptophan and lower levels of kynurenine in cord blood of intrauterine growth-restricted babies. Animal experiments have shown cord blood levels of tryptophan to be related to maternal plasma levels,(49) suggesting that cord blood levels may reflect tryptophan utilization earlier in pregnancy. Therefore, monitoring of tryptophan metabolism throughout pregnancy may have utility in tracking the health of the developing fetus. Similarly, lower levels of multiple acylcarnitine species (C4, C6, C8, C10, C12, C14, and C16) were associated with increased birth weight, likely reflecting differences in energy utilization during development. Carnitine is an essential factor in fatty acid metabolism, and its most important known metabolic function is to transport fatty acids into the mitochondria of cells for β -oxidation.(50) The placenta has a high activity of fatty acid oxidation enzymes,(51) and where defects in long-chain fatty acid oxidation are noted, there is a higher frequency of small-for-gestation-age babies.(52) Walsh et al.(53) reported higher levels of acylcarnitines, including dodecanoylcarnitine identified in this study, in cord blood of infants asphyxiated during pregnancy, emphasizing their importance to healthy fetal development. Clemente et al.(54) recently demonstrated in the INMA and ENVIRONAGE cohorts that placental mitochondrial DNA content is associated with birth weight and may mediate the effects of environmental toxicants on birth weight. This supports our findings of the importance of metabolic pathways related to mitochondrial function. Mitochondria are particularly susceptible to oxidative stress and therefore may play a key role linking the fetal environment to growth.

The ω -3 fatty acid DHA has long been thought to be beneficial to fetal growth, and recent randomized control trials have found that taking DHA supplements by pregnant women was associated with increases in gestational length and birth weight.(55, 56) Here we observed a seemingly paradoxical negative association between cord blood DHA levels and birth weight. However, it has been reported that DHA intake by the mother explains only a small proportion of variance in DHA levels in cord blood,(57) with the rest presumably explained by endogenous processes. Reduced DHA levels in cord blood would result from increased utilization in central nervous tissues and from greater eicosanoid metabolism. We found DHA levels to be associated with a cluster of metabolites tentatively assigned as diacylglycerols, which are molecules required for the production of arachidonic acid, an ω -6 fatty acid also involved in eicosanoid metabolism. Together these results highlight the role of long-chain fatty acid and potentially eicosanoid metabolism in fetal growth. Eicosanoids, in particular, prostaglandins, are produced throughout pregnancy and play a role in regulating the maternal cardiovascular system and, like progesterone, are involved in signaling the onset of labor. Glycerophospholipids also showed significant disruption, indicating a variety of processes occurring in association with these molecular species. The lysoPCs were positively associated with birth weight and were negatively correlated with DHA and the diacylglycerols, suggesting some metabolic dependency. Positive associations between birth weight and lysoPCs have also been recently reported in a targeted analysis.(17) On the contrary, the PCs were associated with lower birth weight. This could reflect some cell membrane damage leading to the subsequent release of phospholipids, which have a variety of different proposed biological properties.(58) There is also recent evidence of antiphospholipid antibodies (and complement activation) cooperating in triggering a local inflammatory process,(59) which may be linked to suboptimal fetal development.

We have complemented classical laboratory-based metabolic feature annotation in this study with the use of the recently developed *Mummichog* algorithm to extract additional biological information at the pathway level. Because the method matches metabolic features to pathways based on annotation through exact mass only and does not account for any other physio-chemical identifiers, results of these analyses

should be interpreted with caution. However, because feature misidentification likely applies equally to both the numerator (significant features) and denominator (total features detected) in enrichment analyses, the impact of wrong annotation may be less dramatic at the pathway level. We found that the majority of pathways identified, including tryptophan metabolism, carnitine shuttle, glycerophospholipid metabolism, and C-21 steroid hormone biosynthesis, were supported by the laboratory annotation of metabolites.

Recent evidence indicates an effect of birth weight on later metabolic profile,(60) with similarities to profiles associated with weight status measured in later life stages, suggesting that the metabolic profile of an individual at the start of life may persist into later life, with implications for health throughout the life course. Therefore, one may speculate that metabolic pathways identified in this study may provide a link between the observed associations with between birth weight and disease in adult life. In particular, in utero levels of progesterone could plausibly mediate the observed association between high birth weight and later development of breast cancer,(6) as evidence is emerging that progesterone in adult life has a protective effect on breast cancer.(61) Conversely, tryptophan levels may play a role in the association between lower birth weight and increased risk of cardio-metabolic disease in later life because tryptophan has been found to be predictive of subsequent development of type II diabetes in adult cohorts.(62) Following up current birth cohorts to adult life would be of great value to investigate these questions.

The main limitation of this study was related to its use of cord blood. Because samples were by necessity collected at the time of delivery, the study was cross-sectional in nature; therefore, it was difficult to disentangle whether perturbed metabolites were a cause or a result of variance in birth weight. However, cord blood provides a window into the direct supply of nutrients and other essential molecules to the developing fetus, and it also provides a snapshot of metabolism at the start of life. Our study was limited in scope to investigate changes specifically associated with low-birth-weight (<2500 g) babies because we sampled from across the general population of births. However, the generalizability of our results, also considering the large number of samples collected from birth cohorts from across Europe, is a strength. Although any single metabolomics method cannot cover all molecules of the cord blood metabolome, the UHPLC–MS platform we used represents a highly sensitive analytical technique able to measure hundreds to thousands of metabolites. Future work may include the incorporation of complementary metabolomics methods, other ‘omic approaches such as DNA methylation analysis and multiplex analysis of cytokines, which are of interest due to their role in mediating hormonal signaling and exploring the role of the cord blood metabolome in postnatal growth and development.

Conclusions

We have described metabolic profiles associated with birth weight among normal deliveries, highlighting the role of multiple metabolites in various pathways including tryptophan metabolism, fatty acid, and glycerophospholipid metabolism, and hormonal signaling. These results will have implications for antenatal and perinatal care, improving understanding of the pathways through which fetal growth may be affected, and may have implications for health in later life.

References

1. Seeds, J. W.; Peng, T. Impaired growth and risk of fetal death: is the tenth percentile the appropriate standard? *Am. J. Obstet. Gynecol.* 1998, 178 (4) 658– 69 DOI: 10.1016/S0002-9378(98)70475-2
2. Ong, K. K.; Ahmed, M. L.; Emmett, P. M.; Preece, M. A.; Dunger, D. B. Association between postnatal catch-up growth and obesity in childhood: prospective cohort study *BMJ* 2000, 320 (7240) 967– 71 DOI: 10.1136/bmj.320.7240.967
3. Goldenberg, R. L.; DuBard, M. B.; Cliver, S. P.; Nelson, K. G.; Blanksona, K.; Ramey, S. L.; Herman, A. Pregnancy outcome and intelligence at age five years *Am. J. Obstet. Gynecol.* 1996, 175 (6) 1511– 5 DOI: 10.1016/S0002-9378(96)70099-6
4. Barker, D. J.; Osmond, C.; Kajantie, E.; Eriksson, J. G. Growth and chronic disease: findings in the Helsinki Birth Cohort *Ann. Hum. Biol.* 2009, 36 (5) 445– 58 DOI: 10.1080/03014460902980295
5. Nilsen, T. I.; Romundstad, P. R.; Troisi, R.; Potischman, N.; Vatten, L. J. Birth size and colorectal cancer risk: a prospective population based study *Gut* 2005, 54 (12) 1728– 32 DOI: 10.1136/gut.2004.060475
6. Ahlgren, M.; Melbye, M.; Wohlfahrt, J.; Sorensen, T. I. Growth patterns and the risk of breast cancer in women *N. Engl. J. Med.* 2004, 351 (16) 1619– 26 DOI: 10.1056/NEJMoa040576
7. Iniguez, C.; Ballester, F.; Costa, O.; Murcia, M.; Souto, A.; Santa-Marina, L.; Aurrekoetxea, J. J.; Espada, M.; Vrijheid, M.; Alvarez-Avellon, S. M.; Alvarez-Pedrerol, M.; Rebagliato, M. Maternal smoking during pregnancy and fetal biometry: the INMA Mother and Child Cohort Study *Am. J. Epidemiol.* 2013, 178 (7) 1067– 75 DOI: 10.1093/aje/kwt085
8. Pedersen, M.; Giorgis-Allemand, L.; Bernard, C.; Aguilera, I.; Andersen, A. M. N.; Ballester, F.; Beelen, R. M. J.; Chatzi, L.; Cirach, M.; Danileviciute, A.; Dedele, A.; Eijdsen, M. V.; Estarlich, M.; Fernández-Somoano, A.; Fernández, M. F.; Forastiere, F.; Gehring, U.; Grazuleviciene, R.; Gruziova, O.; Heude, B.; Hoek, G.; Hoogh, K. D.; van den Hooven, E. H.; Håberg, S. E.; Jaddoe, V. W. V.; Klümper, C.; Korek, M.; Krämer, U.; Lerchundi, A.; Lepeule, J.; Nafstad, P.; Nystad, W.; Patelarou, E.; Porta, D.; Postma, D.; Raaschou-Nielsen, O.; Rudnai, P.; Sunyer, J.; Stephanou, E.; Sørensen, M.; Thiering, E.; Tuffnell, D.; Varró, M. J.; Vrijkotte, T. G. M.; Wijga, A.; Wilhelm, M.; Wright, J.; Nieuwenhuijsen, M. J.; Pershagen, G.; Brunekreef, B.; Kogevinas, M.; Slama, R. Ambient air pollution and low birth weight: A European cohort study (ESCAPE) *Lancet Respir. Med.* 2013, 1, 695– 704 DOI: 10.1016/S2213-2600(13)70192-9
9. Govarts, E.; Nieuwenhuijsen, M.; Schoeters, G.; Ballester, F.; Bloemen, K.; de Boer, M.; Chevrier, C.; Eggesbo, M.; Guxens, M.; Kramer, U.; Legler, J.; Martinez, D.; Palkovicova, L.; Patelarou, E.; Ranft, U.; Rautio, A.; Petersen, M. S.; Slama, R.; Stigum, H.; Toft, G.; Trnovec, T.; Vandentorren, S.; Weihe, P.; Kuperus, N. W.; Wilhelm, M.; Wittsiepe, J.; Bonde, J. P. Birth weight and prenatal exposure to polychlorinated biphenyls (PCBs) and dichlorodiphenyldichloroethylene (DDE): a meta-analysis within 12 European Birth Cohorts *Environ. Health Perspect.* 2011, 120 (2) 162– 70 DOI: 10.1289/ehp.1103767
10. Fanos, V.; Atzori, L.; Makarenko, K.; Melis, G. B.; Ferrazzi, E. Metabolomics application in maternal-fetal medicine *BioMed Res. Int.* 2013, 2013, 1– 9 DOI: 10.1155/2013/720514
11. Horgan, R. P.; Broadhurst, D. I.; Walsh, S. K.; Dunn, W. B.; Brown, M.; Roberts, C. T.; North, R. A.; McCowan, L. M.; Kell, D. B.; Baker, P. N.; Kenny, L. C. Metabolic Profiling Uncovers a

- Phenotypic Signature of Small for Gestational Age in Early Pregnancy *J. Proteome Res.* 2011, 10 (8) 3660– 3673 DOI: 10.1021/pr2002897
12. Maitre, L.; Fthenou, E.; Athersuch, T.; Coen, M.; Toledano, M. B.; Holmes, E.; Kogevinas, M.; Chatzi, L.; Keun, H. C. Urinary metabolic profiles in early pregnancy are associated with preterm birth and fetal growth restriction in the Rhea mother-child cohort study *BMC Med.* 2014, 12, 110 DOI: 10.1186/1741-7015-12-110
 13. Maitre, L.; Villanueva, C. M.; Lewis, M. R.; Ibarluzea, J.; Santa-Marina, L.; Vrijheid, M.; Sunyer, J.; Coen, M.; Toledano, M. B. Maternal urinary metabolic signatures of fetal growth and associated clinical and environmental factors in the INMA study *BMC Med.* 2016, 14, 177 DOI: 10.1186/s12916-016-0706-3
 14. Dessì, A.; Atzori, L.; Noto, A.; Adriaan Visser, G. H.; Gazzolo, D.; Zanardo, V.; Barberini, L.; Puddu, M.; Ottonello, G.; Atzei, A.; Magistris, A. D.; Lussu, M.; Murgia, F.; Fanos, V. Metabolomics in newborns with intrauterine growth retardation (IUGR): urine reveals markers of metabolic syndrome *J. Matern.-Fetal Neonat. Med.* 2011, 24 (sup2) 35– 39 DOI: 10.3109/14767058.2011.605868
 15. Ivorra, C.; García-Vicent, C.; Chaves, F. J.; Monleón, D.; Morales, J. M.; Lurbe, E. Metabolomic profiling in blood from umbilical cords of low birth weight newborns *J. Transl. Med.* 2012, 10, 142 DOI: 10.1186/1479-5876-10-142
 16. Tea, I.; Le Gall, G.; Kuster, A.; Guignard, N.; Alexandre-Gouabau, M.-C.; Darmaun, D.; Robins, R. J. 1H-NMR-based metabolic profiling of maternal and umbilical cord blood indicates altered materno-foetal nutrient exchange in preterm infants *PLoS One* 2012, 7 (1) e29947 DOI: 10.1371/journal.pone.0029947
 17. Hellmuth, C.; Uhl, O.; Standl, M.; Demmelmaier, H.; Heinrich, J.; Koletzko, B.; Thiering, E. Cord Blood Metabolome Is Highly Associated with Birth Weight, but Less Predictive for Later Weight Development *Obes. Facts* 2017, 10 (2) 85– 100 DOI: 10.1159/000453001
 18. Vineis, P.; Chadeau-Hyam, M.; Gmuender, H.; Gulliver, J.; Herceg, Z.; Kleinjans, J.; Kogevinas, M.; Kyrtopoulos, S.; Nieuwenhuijsen, M.; Phillips, D. H.; Probst-Hensch, N.; Scalbert, A.; Vermeulen, R.; Wild, C. P. The exposome in practice: Design of the EXPOsOMICS project *Int. J. Hyg. Environ. Health* 2017, 220 (2) 142– 151 DOI: 10.1016/j.ijheh.2016.08.001
 19. Janssen, B. G.; Madlhoum, N.; Gyselaers, W.; Bijnsens, E.; Clemente, D. B.; Cox, B.; Hogervorst, J.; Luyten, L.; Martens, D. S.; Peusens, M.; Plusquin, M.; Provost, E. B.; Roels, H. A.; Saenen, N. D.; Tsamou, M.; Vriens, A.; Winckelmans, E.; Vrijens, K.; Nawrot, T. S. Cohort Profile: The ENVIRonmental influence ON early AGEing (ENVIRONAGE): a birth cohort study *International journal of epidemiology* 2017, 46 (5) 1386– 1387 DOI: 10.1093/ije/dyw269
 20. Farchi, S.; Forastiere, F.; Vecchi Brumatti, L.; Alviti, S.; Arnofi, A.; Bernardini, T.; Bin, M.; Brescianini, S.; Colelli, V.; Cotichini, R.; Culasso, M.; De Bartolo, P.; Felice, L.; Fiano, V.; Fioritto, A.; Frizzi, A.; Gagliardi, L.; Giorgi, G.; Grasso, C.; La Rosa, F.; Loganes, C.; Lorusso, P.; Martini, V.; Merletti, F.; Medda, E.; Montelatici, V.; Mugelli, I.; Narduzzi, S.; Nistico, L.; Penna, L.; Piscianz, E.; Piscicelli, C.; Poggesi, G.; Porta, D.; Ranieli, A.; Rapisardi, G.; Rasulo, A.; Richiardi, L.; Rusconi, F.; Serino, L.; Stazi, M. A.; Toccaceli, V.; Todros, T.; Tognin, V.; Trevisan, M.; Valencic, E.; Volpi, P.; Ziroli, V.; Ronfani, L.; Di Lallo, D. Piccolipiu, a multicenter birth cohort in Italy: protocol of the study *BMC Pediatr.* 2014, 14, 36 DOI: 10.1186/1471-2431-14-36
 21. Guxens, M.; Ballester, F.; Espada, M.; Fernandez, M. F.; Grimalt, J. O.; Ibarluzea, J.; Olea, N.; Rebagliato, M.; Tardon, A.; Torrent, M.; Vioque, J.; Vrijheid, M.; Sunyer, J. Cohort Profile:

- the INMA--Infancia y Medio Ambiente--(Environment and Childhood) Project International journal of epidemiology 2012, 41 (4) 930– 40 DOI: 10.1093/ije/dyr054
22. Chatzi, L.; Plana, E.; Daraki, V.; Karakosta, P.; Alegkakis, D.; Tsatsanis, C.; Kafatos, A.; Koutis, A.; Kogevinas, M. Metabolic syndrome in early pregnancy and risk of preterm birth *Am. J. Epidemiol.* 2009, 170 (7) 829– 36 DOI: 10.1093/aje/kwp211
 23. Wang, M.; Beelen, R.; Basagana, X.; Becker, T.; Cesaroni, G.; de Hoogh, K.; Dedele, A.; Declercq, C.; Dimakopoulou, K.; Eeftens, M.; Forastiere, F.; Galassi, C.; Grazuleviciene, R.; Hoffmann, B.; Heinrich, J.; Iakovides, M.; Kunzli, N.; Korek, M.; Lindley, S.; Molter, A.; Mosler, G.; Madsen, C.; Nieuwenhuijsen, M.; Phuleria, H.; Pedeli, X.; Raaschou-Nielsen, O.; Ranzi, A.; Stephanou, E.; Sugiri, D.; Stempfelet, M.; Tsai, M. Y.; Lanki, T.; Udvardy, O.; Varro, M. J.; Wolf, K.; Weinmayr, G.; Yli-Tuomi, T.; Hoek, G.; Brunekreef, B. Evaluation of land use regression models for NO₂ and particulate matter in 20 European study areas: the ESCAPE project *Environ. Sci. Technol.* 2013, 47 (9) 4357– 64 DOI: 10.1021/es305129t
 24. Lazar, C. imputeLCMD: A Collection of Methods for Left-Censored Missing Data Imputation, R package, 2.0, 2015. <https://CRAN.R-project.org/package=imputeLCMD>.
 25. Campanella, G. omics: “-omics” Data Analysis Toolbox, R package version 0.1–4.; 2016. <https://CRAN.R-project.org/package=omics>.
 26. R Core Team. R: A Language and Environment for Statistical Computing; Vienna, Austria, 2016. <https://www.R-project.org/>.
 27. Wishart, D. S.; Jewison, T.; Guo, A. C.; Wilson, M.; Knox, C.; Liu, Y.; Djombou, Y.; Mandal, R.; Aziat, F.; Dong, E.; Bouatra, S.; Sinelnikov, I.; Arndt, D.; Xia, J.; Liu, P.; Yallou, F.; Bjorn Dahl, T.; Perez-Pineiro, R.; Eisner, R.; Allen, F.; Neveu, V.; Greiner, R.; Scalbert, A. HMDB 3.0--The Human Metabolome Database in 2013 *Nucleic Acids Res.* 2012, 41 (Database issue) D801– 7 DOI: 10.1093/nar/gks1065
 28. Smith, C. A.; O’Maille, G.; Want, E. J.; Qin, C.; Trauger, S. A.; Brandon, T. R.; Custodio, D. E.; Abagyan, R.; Siuzdak, G. METLIN: a metabolite mass spectral database *Ther. Drug Monit.* 2005, 27 (6) 747– 51 DOI: 10.1097/01.ftd.0000179845.53213.39
 29. Li, L.; Li, R.; Zhou, J.; Zuniga, A.; Stanislaus, A. E.; Wu, Y.; Huan, T.; Zheng, J.; Shi, Y.; Wishart, D. S.; Lin, G. MyCompoundID: Using an Evidence-Based Metabolome Library for Metabolite Identification *Anal. Chem.* 2013, 85 (6) 3401– 3408 DOI: 10.1021/ac400099b
 30. Sumner, L. W.; Amberg, A.; Barrett, D.; Beale, M. H.; Beger, R.; Daykin, C. A.; Fan, T. W. M.; Fiehn, O.; Goodacre, R.; Griffin, J. L.; Hankemeier, T.; Hardy, N.; Harnly, J.; Higashi, R.; Kopka, J.; Lane, A. N.; Lindon, J. C.; Marriott, P.; Nicholls, A. W.; Reilly, M. D.; Thaden, J. J.; Viant, M. R. Proposed minimum reporting standards for chemical analysis: Chemical Analysis Working Group (CAWG) Metabolomics Standards Initiative (MSI) *Metabolomics* 2007, 3, 211– 221 DOI: 10.1007/s11306-007-0082-2
 31. Li, S.; Park, Y.; Duraisingham, S.; Strobel, F. H.; Khan, N.; Soltow, Q. A.; Jones, D. P.; Pulendran, B. Predicting Network Activity from High Throughput Metabolomics *PLoS Comput. Biol.* 2013, 9 (7) e1003123 DOI: 10.1371/journal.pcbi.1003123
 32. Dubois, L.; Ohm Kyvik, K.; Girard, M.; Tatone-Tokuda, F.; Pérusse, D.; Hjelmborg, J.; Skytthe, A.; Rasmussen, F.; Wright, M. J.; Lichtenstein, P.; Martin, N. G. Genetic and environmental contributions to weight, height, and bmi from birth to 19 years of age: An international study of over 12,000 twin pairs *PLoS One* 2012, 7, e30153 DOI: 10.1371/journal.pone.0030153
 33. Masters, E. T.; Jedrychowski, W.; Schleicher, R. L.; Tsai, W. Y.; Tu, Y. H.; Camann, D.; Tang, D.; Perera, F. P. Relation between prenatal lipid-soluble micronutrient status,

- environmental pollutant exposure, and birth outcomes *Am. J. Clin. Nutr.* 2007, 86 (4) 1139–45
34. Rondo, P. H.; Abbott, R.; Tomkins, A. M. Vitamin A and neonatal anthropometry *Journal of tropical pediatrics* 2001, 47 (5) 307– 10 DOI: 10.1093/tropej/47.5.307
 35. Gazala, E.; Sarov, B.; HersHKovitz, E.; Edvardson, S.; Sklan, D.; Katz, M.; Friger, M.; Gorodischer, R. Retinol concentration in maternal and cord serum: its relation to birth weight in healthy mother-infant pairs *Early Hum. Dev.* 2003, 71 (1) 19– 28 DOI: 10.1016/S0378-3782(02)00096-8
 36. Ghebremeskel, K.; Burns, L.; Burden, T. J.; Harbige, L.; Costeloe, K.; Powell, J. J.; Crawford, M. Vitamin A and related essential nutrients in cord blood: relationships with anthropometric measurements at birth *Early Hum. Dev.* 1994, 39 (3) 177– 88 DOI: 10.1016/0378-3782(94)90196-1
 37. Clagett-Dame, M.; DeLuca, H. F. The role of vitamin A in mammalian reproduction and embryonic development *Annu. Rev. Nutr.* 2002, 22, 347– 81 DOI: 10.1146/annurev.nutr.22.010402.102745E
 38. Chelchowska, M.; Ambroszkiewicz, J.; Gajewska, J.; Laskowska-Klita, T.; Leibschang, J. The effect of tobacco smoking during pregnancy on plasma oxidant and antioxidant status in mother and newborn *Eur. J. Obstet. Gynecol. Reprod. Biol.* 2011, 155 (2) 132– 6 DOI: 10.1016/j.ejogrb.2010.12.006
 39. Li, T.; Molteni, A.; Latkovich, P.; Castellani, W.; Baybutt, R. C. Vitamin A Depletion Induced by Cigarette Smoke Is Associated with the Development of Emphysema in Rats *J. Nutr.* 2003, 133 (8) 2629– 2634 DOI: 10.1093/jn/133.8.2629
 40. Lagiou, P.; Samoli, E.; Hsieh, C. C.; Lagiou, A.; Xu, B.; Yu, G. P.; Onoyama, S.; Chie, L.; Adami, H. O.; Vatten, L. J.; Trichopoulos, D.; Williams, M. A. Maternal and cord blood hormones in relation to birth size *Eur. J. Epidemiol.* 2014, 29, 343– 351 DOI: 10.1007/s10654-014-9914-3
 41. Hartwig, I. R.; Pincus, M. K.; Diemert, A.; Hecher, K.; Arck, P. C. Sex-specific effect of first-trimester maternal progesterone on birthweight *Hum. Reprod.* 2013, 28 (1) 77– 86 DOI: 10.1093/humrep/des367
 42. He, S.; Allen, J. C., Jr.; Malhotra, R.; Ostbye, T.; Tan, T. C. Association of maternal serum progesterone in early pregnancy with low birth weight and other adverse pregnancy outcomes *J. Matern.-Fetal Neonat. Med.* 2016, 29 (12) 1999– 2004 DOI: 10.3109/14767058.2015.1072159
 43. Mucci, L. A.; Lagiou, P.; Tamimi, R. M.; Hsieh, C. C.; Adami, H. O.; Trichopoulos, D. Pregnancy estriol, estradiol, progesterone and prolactin in relation to birth weight and other birth size variables (United States) *Cancer, Causes Control* 2003, 14 (4) 311– 8 DOI: 10.1023/A:1023966813330
 44. Lagiou, P.; Hsieh, C. C.; Samoli, E.; Lagiou, A.; Xu, B.; Yu, G. P.; Onoyama, S.; Chie, L.; Vatten, L. J.; Adami, H. O.; Trichopoulos, D.; Williams, M. A. Associations of placental weight with maternal and cord blood hormones *Annals of epidemiology* 2013, 23 (11) 669– 73 DOI: 10.1016/j.annepidem.2013.08.003
 45. Tal, R.; Taylor, H.; Burney, R.; Endocrinology of Pregnancy [Updated 2015 Dec 7]. In *Endotext* [Internet], De Groot, L.; Chrousos, G.; Dungan, K., Eds.; MDText.com, Inc.: South Dartmouth, MA, 2000. <https://www-ncbi-nlm-nih.gov/bibliopass.unito.it/books/NBK278962/>.
 46. Badawy, A. A. Effects of pregnancy on tryptophan metabolism and disposition in the rat *Biochem. J.* 1988, 255 (1) 369– 72

47. Badawy, A. a.-B. The tryptophan utilization concept in pregnancy *Obstetrics & gynecology science* 2014, 57, 249– 59 DOI: 10.5468/ogs.2014.57.4.249
48. Favretto, D.; Cosmi, E.; Ragazzi, E.; Visentin, S.; Tucci, M.; Fais, P.; Cecchetto, G.; Zanardo, V.; Viel, G.; Ferrara, S. D. Cord blood metabolomic profiling in intrauterine growth restriction *Anal. Bioanal. Chem.* 2012, 402, 1109– 1121 DOI: 10.1007/s00216-011-5540-z
49. Tsuji, A.; Nakata, C.; Sano, M.; Fukuwatari, T.; Shibata, K. L-Tryptophan Metabolism in Pregnant Mice Fed a High L-Tryptophan Diet and the Effect on Maternal, Placental, and Fetal Growth *Int. J. Tryptophan Res.* 2013, 6, 21– 33 DOI: 10.4137/IJTR.S12715
50. Borum, P. R. Carnitine in neonatal nutrition *Journal of child neurology* 1995, 10 (Suppl 2) 2S25– 2S31 DOI: 10.1177/0883073895010002S04
51. Oey, N. A.; den Boer, M. E.; Ruiter, J. P.; Wanders, R. J.; Duran, M.; Waterham, H. R.; Boer, K.; van der Post, J. A.; Wijburg, F. A. High activity of fatty acid oxidation enzymes in human placenta: implications for fetal-maternal disease *J. Inherited Metab. Dis.* 2003, 26 (4) 385– 92 DOI: 10.1023/A:1025163204165
52. Tyni, T.; Ekholm, E.; Pihko, H. Pregnancy complications are frequent in long-chain 3-hydroxyacyl-coenzyme A dehydrogenase deficiency *Am. J. Obstet. Gynecol.* 1998, 178 (3) 603– 8 DOI: 10.1016/S0002-9378(98)70446-6
53. Walsh, B. H.; Broadhurst, D. I.; Mandal, R.; Wishart, D. S.; Boylan, G. B.; Kenny, L. C.; Murray, D. M. The metabolomic profile of umbilical cord blood in neonatal hypoxic ischaemic encephalopathy *PLoS One* 2012, 7 (12) e50520 DOI: 10.1371/journal.pone.0050520
54. Clemente, D. B.; Casas, M.; Vilahur, N.; Begiristain, H.; Bustamante, M.; Carsin, A. E.; Fernandez, M. F.; Fierens, F.; Gyselaers, W.; Iniguez, C.; Janssen, B. G.; Lefebvre, W.; Llop, S.; Olea, N.; Pedersen, M.; Pieters, N.; Santa Marina, L.; Souto, A.; Tardon, A.; Vanpoucke, C.; Vrijheid, M.; Sunyer, J.; Nawrot, T. S. Prenatal Ambient Air Pollution, Placental Mitochondrial DNA Content, and Birth Weight in the INMA (Spain) and ENVIRONAGE (Belgium) Birth Cohorts *Environ. Health Perspect.* 2016, 124 (5) 659– 65 DOI: 10.1289/ehp.1408981
55. Carlson, S. E.; Colombo, J.; Gajewski, B. J.; Gustafson, K. M.; Mundy, D.; Yeast, J.; Georgieff, M. K.; Markley, L. A.; Kerling, E. H.; Shaddy, D. J. DHA supplementation and pregnancy outcomes *Am. J. Clin. Nutr.* 2013, 97 (4) 808– 15 DOI: 10.3945/ajcn.112.050021
56. Harris, M. A.; Reece, M. S.; McGregor, J. A.; Wilson, J. W.; Burke, S. M.; Wheeler, M.; Anderson, J. E.; Auld, G. W.; French, J. I.; Allen, K. G. The Effect of Omega-3 Docosahexaenoic Acid Supplementation on Gestational Length: Randomized Trial of Supplementation Compared to Nutrition Education for Increasing n-3 Intake from Foods *BioMed Res. Int.* 2015, 2015, 1– 8 DOI: 10.1155/2015/123078
57. Muhlhausler, B. S.; Gibson, R. A.; Yelland, L. N.; Makrides, M. Heterogeneity in cord blood DHA concentration: towards an explanation *Prostaglandins, Leukotrienes Essent. Fatty Acids* 2014, 91 (4) 135– 40 DOI: 10.1016/j.plefa.2014.07.013
58. Fruhwirth, G. O.; Loidl, A.; Hermetter, A. Oxidized phospholipids: from molecular properties to disease *Biochim. Biophys. Acta, Mol. Basis Dis.* 2007, 1772 (7) 718– 36 DOI: 10.1016/j.bbadis.2007.04.009
59. Tincani, A.; Cavazzana, I.; Ziglioli, T.; Lojacono, A.; De Angelis, V.; Meroni, P. Complement activation and pregnancy failure *Clin. Rev. Allergy Immunol.* 2010, 39 (3) 153– 9 DOI: 10.1007/s12016-009-8183-5

60. Würtz, P.; Wang, Q.; Niironen, M.; Tynkkynen, T.; Tiainen, M.; Drenos, F.; Kangas, A. J.; Soininen, P.; Skilton, M. R.; Heikkilä, K.; Pouta, A.; Kähönen, M.; Lehtimäki, T.; Rose, R. J.; Kajantie, E.; Perola, M.; Kaprio, J.; Eriksson, J. G.; Raitakari, O. T.; Lawlor, D. A.; Davey Smith, G.; Järvelin, M.-R.; Ala-Korpela, M.; Auro, K. Metabolic signatures of birthweight in 18 288 adolescents and adults *International journal of epidemiology* 2016, 45, 1539– 1550 DOI: 10.1093/ije/dyw255
61. Asi, N.; Mohammed, K.; Haydour, Q.; Gionfriddo, M. R.; Vargas, O. L.; Prokop, L. J.; Faubion, S. S.; Murad, M. H. Progesterone vs. synthetic progestins and the risk of breast cancer: a systematic review and meta-analysis *Syst. Rev.* 2016, 5 (1) 121 DOI: 10.1186/s13643-016-0294-5
62. Chen, T.; Zheng, X.; Ma, X.; Bao, Y.; Ni, Y.; Hu, C.; Rajani, C.; Huang, F.; Zhao, A.; Jia, W.; Jia, W. Tryptophan Predicts the Risk for Future Type 2 Diabetes *PLoS One* 2016, 11 (9) e0162192 DOI: 10.1371/journal.pone.0162192